Effect of Salicylic Acid on Physico-chemical Attributes and Shelf Life of Tomato Fruits at Refrigerated Storage

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Abstract

Tomato, the frequently consumed solanaceous fruit vegetable faced rapid deterioration of fruit edible quality under ambient storage and though refrigerated storage has proved better retention but mostly with the outbreak of chilling injury. Therefore, attempt with salicylic acid, a safe phenolic having identified anti-ethylene effect, has been made during this study to evaluate its effect on shelf life and quality attributes of tomato fruit in refrigerated storage. Fruits of tomato variety Samrudhi at pink to light red stage were treated with different concentration of salicylic acid (0.2−1.2 mM) along with control (water dipped) and were kept at 4−5 °C with 65−80% relative humidity in four replications following complete randomized design. Results showed salicylic acid (SA) treated fruits had better keeping quality than control though in concentration dependant manner. At 21 days after storage (DAS), fruits kept in control condition lost 13.69% of weight compared with the fruits treated with SA at 1.2 mM (5.79%). SA at 1-1.2 mM significantly reduced chilling injury (Chilling Injury Index/CII: 1.44−2.88) compared with fruits at control (CII: 4.98). Fruits at control showed deterioration in sugar content along with acidity and ascorbic acid. Further, it took rapid increase in pigment whereas, SA treated fruit in relatively higher concentration (0.8−1.2 mM), maintained fruit quality attributes with consistent intensification of carotenoids and lycopene even at 21 DAS. SA at 1.2 mM recorded maximum shelf life (32.75 days) to be considered as the best treatment having high maintenance of fruit edible qualities.

Keywords

Refrigerated storage, salicylic acid, chilling injury, lycopene, shelf life, tomato

1. Introduction

Tomato is an important warm season fruit vegetable grown throughout the world and occupying significant position in Indian vegetable production (Gopalakrishnan, 2007). Tomato belongs to the family Solanaceae, genus Lycopersicon and sub family Solanoidae. Tomato fruits are rich source of minerals like calcium, sodium, copper and other trace elements, vitamins like vitamin A (900 IU), vitamin C (27 mg), vitamin B complex along with organic acids like citric, formic and acetic acid and essential amino acids for which tomato occupies a important position in the list of protective foods (Gopalakrishnan, 2007). Tomato is known to have an anti-cancer property due to presence of the red colour pigment lycopene (Rao and Agarwal, 1999). India is the 2nd highest producer of tomato (182.26 lakh MT) next to China (Anon., 2014a). Tomato is exported mainly to the countries like Pakistan, UAE, Bangladesh, Nepal, Saudi Arabia, Oman, Maldives, Bahrain, Russia and Malawi from India and the fresh export of tomato is 3.44 lakh t worth 55305 lakhs, which is quite less (Anonymous, 2014b).

Tomato is regarded as the most common and frequently used fruit vegetable which always having a good market demand. However, it encounters several problems in the transportation, storage and marketing since it is highly perishable (Pila et al., 2010). Due to this, it has been resulted in the loss during harvesting, transportation and consumption of fresh tomatoes and which accounts to 20−25% loss in tropical countries (Aworth and Olorunda, 1981). Apart from loss caused through transport and marketing; cold storage, perhaps enhanced shelf life, however, with chilling injury. Salicylic acid is reported as a natural and safe phenolic compound which has been found to generate a wide range of metabolic and physiological responses in plants and act as potential bio agent in controlling post harvest loss of horticultural crops and delay in ripening through inhibition of ethylene biosynthesis or action (Asghari and Aghdam, 2010). Salicylic acid has potential reports for alleviating chilling injury in sweet orange (Ahmad et al., 2013) and strawberry (Babalar et al., 2007) and extending shelf life by maintaining physico-chemical attributes both in cold or at ambient storage (Barman and Asrey, 2014; Mandal et al.,...
2016). Therefore, the present investigation was taken up to study the effect of salicylic acid in a view to extend shelf life and maintain the quality of tomato.

2. Materials and Methods

2.1. Location of experiment

The experiment was carried out during May–June, 2015, at Research Laboratory, Department of Horticulture, Aromatic & Medicinal Plants, Mizoram University; with freshly harvested fruits of tomato cv. Samrudhi (F, Hybrid from East-West Seed International Ltd., Thailand) obtained from a local tomato grower of Durtlang village, Aizawl district of Mizoram. Fruits cultivated under green house condition were selected as samples with specific maturity indices i.e. pink to light red stage.

2.2. Treatments

Seven post-harvest treatments viz. fruit dipping in Salicylic Acid (SA) at 0.2 mM l\(^{-1}\), 0.4 mM l\(^{-1}\), 0.6 mM l\(^{-1}\), 0.8 mM l\(^{-1}\), 1.0 mM l\(^{-1}\), 1.2 mM l\(^{-1}\) and control (treated with water: T\(_0\)) with four replications were used and statistical analysis was done by following complete randomized design (Gomez and Gomez, 1984). Salicylic Acid of respective concentrations was prepared by diluting in warm distilled water and cooled down before dipping of fruit for 5 minutes whereas fruit samples dipped in distilled water for 5 minutes were taken as control. The entire experiment was conducted at refrigerated condition (4–5 °C with 65–80% relative humidity).

2.3. Determination of weight loss

Fruits for each treatment were tagged and weighed at 7 days interval using a digital electronic balance. The percentage weight loss was calculated by the following equation:-

\[
\text{Percentage weight loss at } n^{th} \text{ day} = \frac{\text{wt. loss (0 day-n}^{th} \text{ day)}}{\text{Wt. at 0 day} \times 100}
\]

2.4. Biochemical parameters

The fruits were prepared for analysis by cutting and macerating the flesh with mortar and pestle and strained with clean muslin cloth. Analyses were carried out immediately for total soluble solids (TSS), total sugar, reducing sugar, titratable acidity, TSS: acid ratio and ascorbic acid content.

2.4.1. Determination of total soluble solids (TSS)

The total soluble solids of the fruits were determined with the help of hand refractometer calibrated in °Brix at 20 °C with necessary correction factor.

2.4.2. Determination of total sugar and reducing sugar

The total sugar and reducing sugar content of fruit juice were estimated by standard procedure of A.O.A.C. (1990) using Fehling’s A and Fehling’s B reagents with methylene blue as an indicator through copper reduction method.

2.4.3. Determination of titratable acidity

Total titratable acidity was determined by titrating the extracted juice against N/10 NaOH (sodium hydroxide) using phenolphthalein as indicator and expressed in percentage (A.O.A.C., 1990).

2.4.4. Determination of TSS/acid ratio

The ratio for fruit juice under each treatment was calculated by dividing TSS value by titratable acidity content of fruit.

2.4.5. Determination of ascorbic acid

2, 6 dichlorophenol indophenols dye titration method was used to estimate the ascorbic acid content of fruit (A.O.A.C., 1990; Ranganna, 1977) and expressed as mg 100 g\(^{-1}\) of fruit.

2.5. Determination of lycopene

Determination of lycopene was performed by using acetone, petroleum ether and anhydrous sodium sulphate following the procedure described by Ranganna (1977). Absorbance was measured using a Digital Spectrophotometer at 503 nm. Final value of lycopene content was converted into μg g\(^{-1}\) unit.

2.6. Determination of carotenoids

Carotene content of the fruit was determined by using acetone, hexane and magnesium carbonate following the standard procedure given by Sadasivam and Manickam (1997). Determination was done by calculating carotene (mg 100 g\(^{-1}\)) in the sample using standard curve prepared with different concentration of β-carotene standard and measuring absorbance at 436 nm wave length using a digital spectrophotometer. Final value of carotene content was converted into μg g\(^{-1}\) unit.

2.7. Chilling injury index (CII)

Chilling injury index based on external damage was scored on each fruit using a subjective scale: = no damage, 2 = slight damage, 3 = medium damage, 4 = severe damage, 5 = very severe damage. The CII was calculated on 21 days after storage (DAS) according to the formula used by Mohammed and Wickham (1996).

\[
\text{CII index} = \frac{\sum_{0}^{5} \text{Injury level} \times \text{No. of fruits at this level}}{\text{Total no. of fruits}}
\]

2.8. Shelf life of fruit

Optimum shelf life (days) of fruit under different treatment in refrigerated condition were evaluated depending on the visual observation of chilling injury, fruit physico-chemical parameters and counting the days from harvest to the day with maximum edible and marketable quality (Pila et al., 2010; Moneruzzaman et al., 2009; Mandal et al., 2015).

3. Results and Discussion

3.1. Physiological weight loss

Present study manifested that tomato fruit significantly lost its physiological weight during storage at refrigerated condition
in all the treatments. Generally the weight loss of tomato fruits increased progressively during their storage (Pila et al., 2010). Znidarcic and Pozrol (2006) also reported about loss of physiological weight of tomato even in storage under low temperature of 5−10 °C. Result of the present investigation indicated that tomato fruit which were not treated with SA showed considerably high percentage of weight loss (6.64%) after 14 days of storage (Table 1). At 21 DAS, it was the fruit at control which got maximum loss in physiological weight (13.69%). Okolie and Sanni (2012) reported that fruits at control had maximum weight loss (6.87±0.01%) at 14 DAS of cold storage. Gharezi et al. (2012) opined that weight loss of fresh tomatoes is primarily due to transpiration and respiration. Transpiration is a mechanism in which water is lost due to differences in vapour pressure of water in the atmosphere and the transpiring surface. Respiration causes a weight reduction because a carbon atom is lost from the fruit each time a carbon dioxide molecule is produced from an absorbed oxygen molecule and evolved into atmosphere (Bhowmik and Pan, 1992). It was seen that SA treated fruits had significantly less percentage of weight loss during storage. At 21 DAS, SA at 1.2mM caused minimum weight loss (5.79%) followed by the fruits treated with SA at 1.0 mM (6.07%). Tareen et al. (2012) reported that Kiwi fruits treated with SA at 0.2 mM showed lowest loss of fruit weight. SA has been reported to close stomata which results in suppressed respiration rate and minimize weight loss of fruits (Manthe et al., 1992; Zheng and Zhang, 2004). Kamal Kant et al. (2013) reported that exogenous application of SA reduced weight loss of tomato fruits.

3.2. TSS, titrable acidity and TSS:acid ratio

Present study revealed that tomato fruits stored at refrigerated condition had gradual increase in TSS value with decreasing titrable acidity which inturn increased TSS: acid ratio at the end of 21 days storage (Table 2). Gharezi et al. (2012) had similar kind of observation in cold storage of cherry tomato, where TSS value increased upto 10 DAS afterward declined along with decrease in fruit titrable acidity. They reported that fruits at control had 4.5% TSS at 0 DAS, which increased to 4.6% at 10 DAS and declined to 4.5% TSS finally at 15 DAS at cold storage, whereas, titrable acidity was 0.4% at 0 DAS which become 0.35% at 15 DAS. In the present study it was found that fruits at control showed 4.72 °Brix TSS at 7 DAS which increased to 6.08 °Brix at 14 DAS and finally declined to 4.82 °Brix at 21 DAS, whereas titrable acidity was 0.26% at 7 DAS which ultimately reduced to 0.13% at 21 DAS. Fruits treated with SA at 1−1.2 mM showed delayed increase

<table>
<thead>
<tr>
<th>Treatments</th>
<th>PW: Percentage of weight loss (%)</th>
<th>CII: Chilling injury index</th>
<th>Shelf life (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SA at 0.2 mM</td>
<td>4.03 5.95 8.93</td>
<td>4.80 25.25</td>
<td></td>
</tr>
<tr>
<td>SA at 0.2 mM</td>
<td>3.84 5.82 8.61</td>
<td>4.32 26.75</td>
<td></td>
</tr>
<tr>
<td>SA at 0.4 mM</td>
<td>3.78 5.67 7.24</td>
<td>3.60 27.50</td>
<td></td>
</tr>
<tr>
<td>SA at 0.6 mM</td>
<td>3.05 4.67 6.31</td>
<td>3.60 28.75</td>
<td></td>
</tr>
<tr>
<td>SA at 0.8 mM</td>
<td>2.92 4.59 6.07</td>
<td>2.88 30.75</td>
<td></td>
</tr>
<tr>
<td>SA at 1.0 mM</td>
<td>2.75 3.72 5.79</td>
<td>1.44 32.75</td>
<td></td>
</tr>
<tr>
<td>SA at 1.2 mM</td>
<td>5.24 6.64 13.69</td>
<td>4.98 24.00</td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>0.2794 0.2886 0.4714</td>
<td>- 0.9177</td>
<td></td>
</tr>
<tr>
<td>CD at (p=0.05)</td>
<td>0.8218 0.8491 1.3867</td>
<td>- 2.6996</td>
<td></td>
</tr>
</tbody>
</table>

PW: Percentage of weight loss (%); CII: Chilling injury index

<table>
<thead>
<tr>
<th>Treatments</th>
<th>TSS (°Brix)</th>
<th>Titrable acidity (%)</th>
<th>TSS: Acıd ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>SA at 0.2 mM</td>
<td>4.44 5.92 4.92</td>
<td>0.32 0.24 0.19</td>
<td>13.88 24.67 25.89</td>
</tr>
<tr>
<td>SA at 0.2 mM</td>
<td>4.42 5.82 4.96</td>
<td>0.32 0.26 0.19</td>
<td>13.81 22.38 26.11</td>
</tr>
<tr>
<td>SA at 0.4 mM</td>
<td>4.28 5.18 5.04</td>
<td>0.38 0.32 0.22</td>
<td>11.26 16.19 22.91</td>
</tr>
<tr>
<td>SA at 0.6 mM</td>
<td>4.12 4.60 5.16</td>
<td>0.42 0.32 0.26</td>
<td>9.81 14.38 19.85</td>
</tr>
<tr>
<td>SA at 0.8 mM</td>
<td>4.10 4.52 5.28</td>
<td>0.42 0.36 0.28</td>
<td>9.76 12.56 18.86</td>
</tr>
<tr>
<td>SA at 1.0 mM</td>
<td>4.08 4.36 5.68</td>
<td>0.45 0.38 0.32</td>
<td>9.07 11.47 17.75</td>
</tr>
<tr>
<td>SA at 1.2 mM</td>
<td>4.72 6.08 4.82</td>
<td>0.26 0.19 0.13</td>
<td>18.15 32.00 32.38</td>
</tr>
<tr>
<td>Control</td>
<td>0.2480 0.3151 0.2223</td>
<td>0.0385 0.0359 0.0302</td>
<td>1.1902 1.1088 1.3621</td>
</tr>
<tr>
<td>CD (p=0.05)</td>
<td>0.7296 0.9268 0.6539</td>
<td>0.1131 0.1057 0.0889</td>
<td>3.5011 3.2617 4.0066</td>
</tr>
</tbody>
</table>

*: Non significant
in TSS value. Upto 14 DAS, it was found minimum (4.52 and 4.36 °Brix) under these treatments which later on increased (5.28 and 5.68 °Brix) at 21 DAS. Similarly, drop of titrable acidity was considerably slow under these treatments which resulted in low TSS: acid ratio even at 21 DAS. Aghdam et al., 2010 reported that MeSA at 32 µl l\(^{-1}\) maintained a lower TSS content of Kiwi fruits at cold storage. Kamal Kant et al. (2013) found higher titrable acidity of tomato fruits treated with SA at 0.75 mM. Accumulation of TSS and drop of titrable acidity was found slow in SA treated fruits probably due to slowing down of respiration and metabolic activity, hence retarding the ripening process of fruits. Low TSS: acid ratio of fruits treated with SA at 1−1.2 mM signified that these treatments has potential anti ripening effect (Asghari and Aghdam, 2010).

3.3. Total sugar and reducing sugar

Present study showed that fruits at control or treated with SA at lower concentration (0.2−0.8 mM) had accumulation of total sugar upto 14 DAS and afterward reduced at 21 DAS, whereas reducing sugar consistently reduced through the period of storage (Table 3). Fruits treated with SA at 1−1.2 mM caused delayed accumulation of total sugar with slow decline of reducing sugar. Aghdam et al. (2010) proposed that MeSA reduced ethylene production which may result to decrease sucrose-phosphate synthase activity leading to delay in sucrose synthesis.

3.4. Ascorbic acid

Tomato fruits are rich in ascorbic acid. The ascorbic acid content of ripe tomato fruits ranger from 15 mg to 23 mg 100 g\(^{-1}\) (Sanchez-Moreno et al., 2006). Preservation of ascorbic acid during storage is a difficult task since it undergoes oxidation (Cantwell et al., 2009). A decrease of ascorbic acid content of fruits indicate senescence (Sammi and Masud, 2007). In our study, it has been observed that fruit vitamin C content reduced with increase in storage time. Fruits at control recorded minimum ascorbic acid content (11.27 mg 100 g\(^{-1}\)) at 21 DAS (Table 3). Gherezi et al. (2012) also had similar kind of observation, where fruits got consistent decrease in ascorbic acid content and fruits at control showed minimum vitamin C (17.29 mg 100 g\(^{-1}\)) at 14 DAS at cold storage. Fruits treated with SA showed significantly slow drop in ascorbic acid content. SA at 1.2 mM caused maximum retention of ascorbic acid (16.43 mg 100 g\(^{-1}\)) at 21 DAS. Renhua et al. (2008) reported that application of SA was found to be effective in reducing the rate of respiration and ethylene production and maintaining higher amount of ascorbic acid.

### Table 3: Effect of selected post-harvest treatments on total sugar, reducing sugar and ascorbic acid content of tomato fruits under refrigerated storage

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Total sugar (%)</th>
<th>Reducing sugar (%)</th>
<th>Ascorbic acid (mg 100 g(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>7 DAS</td>
<td>14 DAS</td>
<td>21 DAS</td>
</tr>
<tr>
<td>SA at 0.2 mM</td>
<td>4.72</td>
<td>4.86</td>
<td>4.11</td>
</tr>
<tr>
<td>SA at 0.2 mM</td>
<td>4.68</td>
<td>4.70</td>
<td>4.23</td>
</tr>
<tr>
<td>SA at 0.4 mM</td>
<td>4.60</td>
<td>4.68</td>
<td>4.38</td>
</tr>
<tr>
<td>SA at 0.6 mM</td>
<td>4.55</td>
<td>4.65</td>
<td>4.46</td>
</tr>
<tr>
<td>SA at 0.8 mM</td>
<td>4.25</td>
<td>4.36</td>
<td>4.52</td>
</tr>
<tr>
<td>SA at 1.0 mM</td>
<td>3.87</td>
<td>4.21</td>
<td>4.58</td>
</tr>
<tr>
<td>SA at 1.2 mM</td>
<td>5.02</td>
<td>5.42</td>
<td>4.02</td>
</tr>
<tr>
<td>Control</td>
<td>0.1446</td>
<td>0.1746</td>
<td>0.1311</td>
</tr>
<tr>
<td>CD (p=0.05)</td>
<td>0.4255</td>
<td>0.5135</td>
<td>0.3857</td>
</tr>
</tbody>
</table>

3.5. Carotenoids and lycopene

In the present study, it was observed that carotenoids and lycopene content increased considerably during the period of storage. It was found that fruits at control had carotenoids content of 33.56 µg g\(^{-1}\) with 22.43 µg g\(^{-1}\) of lycopene (Table 4). Gherezi et al. (2012) reported that cold stored tomato fruits got consistent increase in lycopene content and was scored maximum (4.913 mg 100 g\(^{-1}\)) at 14 DAS. It was recorded in the present study, that accumulation of carotenoids and lycopene was slow in case of the fruits treated with SA at relatively higher concentration. At 21 DAS, carotenoids and lycopene content was found minimum (21.08 and 12.70 µg g\(^{-1}\)) in case of the fruits treated with SA at 1.2 mM. Ding et al. (2002) observed minimum fruit colour (13.74±10.80) in case of treatment with MeSA at 0.5 mM. Pila et al. (2010) found that tomato fruits had low carotenoids and lycopene accumulation (29.35 and 28.57 µg g\(^{-1}\)) during storage when treated with SA at 0.4 mM.

3.6. Chilling injury

Determination of chilling injury is of utmost important while storing of fruits at refrigerated condition as it is related to fruit quality, marketability, disease incidence and overall shelf life
Table 4: Effect of selected post-harvest treatments on carotenoids and lycopene content of tomato fruits under refrigerated storage

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Carotenoids (µg g⁻¹)</th>
<th>Lycopene (µg g⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>7 DAS</td>
<td>14 DAS</td>
</tr>
<tr>
<td>SA at 0.2 mM</td>
<td>7.60</td>
<td>14.12</td>
</tr>
<tr>
<td>SA at 0.2 mM</td>
<td>7.20</td>
<td>12.64</td>
</tr>
<tr>
<td>SA at 0.4 mM</td>
<td>6.40</td>
<td>10.83</td>
</tr>
<tr>
<td>SA at 0.6 mM</td>
<td>6.04</td>
<td>9.96</td>
</tr>
<tr>
<td>SA at 0.8 mM</td>
<td>5.82</td>
<td>9.80</td>
</tr>
<tr>
<td>SA at 1.0 mM</td>
<td>5.20</td>
<td>6.92</td>
</tr>
<tr>
<td>SA at 1.2 mM</td>
<td>8.40</td>
<td>14.32</td>
</tr>
<tr>
<td>Control</td>
<td>0.2790</td>
<td>0.5225</td>
</tr>
<tr>
<td>CD (p=0.05)</td>
<td>0.8206</td>
<td>1.5369</td>
</tr>
</tbody>
</table>

of the stored fruit. Present study showed that tomato fruits got maximum CII (4.98) at control, whereas CII was recorded considerably low (1.44−2.88) in case of fruits treated with SA at 1−1.2 mM (Plate 1). Ding et al. (2002) reported that tomato fruits got maximum chilling injury at control compared with the fruits treated with MeSA at 0.5 mM. Aghdam et al. (2012) found that maximum CII (>4.5) in case of fruits at control compared with fruits treated with SA at 1.2 mM (Plate 1). It was reported that SA had significant influence on respiration, ethylene biosynthesis, fruit ripening and had potentiality of controlling post harvest fruit decay (Asghari and Aghdam, 2010; Babalar et al., 2007) which may be the reason for having more shelf life under SA treated fruits. Kamal Kant et al. (2013) also found that the shelf life of tomato fruits has been increased for 7 days more than fruits at control when treated with SA at 0.75 mM.

3.7. Shelf life

Present study showed that fruits at control had minimum shelf life (24 days) when stored at refrigerated (4–5 °C) condition. Okolie and Sanni (2012) reported of 28 days of storage of tomato under cold storage. SA treated fruits had better shelf life (ranged between 25.25−32.75 days) than control. Shelf life was found maximum (32.75 days) in case of the fruits treated with SA at 1.2 mM (Plate 1). It was reported that SA had significant influence on respiration, ethylene biosynthesis, fruit ripening and had potentiality of controlling post harvest fruit decay (Asghari and Aghdam, 2010; Babalar et al., 2007) which may be the reason for having more shelf life under SA treated fruits. Kamal Kant et al. (2013) also found that the shelf life of tomato fruits has been increased for 7 days more than fruits at control when treated with SA at 0.75 mM.

4. Conclusion

The results of the present experiment showed that SA at 1.0-1.2 mM maybe the effective post harvest treatment to extend shelf life while maintaining the physico-chemical attributes of tomato cv. Samrudhi under refrigerated storage.

5. References


Rao, A., Agarwal, S., 1999. Role of lycopene as antioxidant